

# Journal of Medicinal and Chemical Sciences

Journal homepage: <a href="http://www.jmchemsci.com/">http://www.jmchemsci.com/</a>

Original Article

# Role of MiRNA126 as a Biomarker for Coronary Artery Disease in Iraqi Patients

# Hibah Rajaa Taher<sup>1,\*</sup>, Perry Habib Saifullah<sup>2</sup>

<sup>1</sup>Ministry of Science and Technology, Department of Materials Research, Baghdad, Iraq <sup>2</sup>Department of Chemistry, College of Science for Women, University of Baghdad, Baghdad, Iraq

#### ARTICLE INFO

#### **Article history**

Receive: 2022-06-24

Received in revised: 2022-08-03

Accepted: 2022-09-11

Manuscript ID: JMCS-2208-1665 Checked for Plagiarism: **Yes** 

Language Editor: Dr. Fatimah Ramezani

Editor who approved publication:

Dr. Nenad Ignjatovic

# DOI:10.26655/JMCHEMSCI.2023.5.22

#### KEYWORDS

Coronary artery disease MicroRNA miR126 Il6,hn

#### ABSTRACT

The current study aimed to evaluate the role and gene target of miR-126 in Iraqi patients with Coronary Artery Disease (CAD) and atherosclerosis (AS). By coverage average age (35 to 65) years old of both genders, a total of 40 patients with CAD and 40 healthy people as a control group were enrolled in this clinical study. The blood samples of patients were taken from Ibn Al-Bitar Diagnostic Center for Heart Surgery in Baghdad/Iraq in December 2020 to April 2021. Overall, the CAD patients had significantly higher (except HDL) total lipid profile (total chol., total Trig., LDL, and VLDL, age, and IL6 levels). Statistically no significant correlation was found between miRNA-126 and lipid parameters in all studied subject. The correlation was found between Triglycerides and his-Troponin (r=-0.521, pv=0.032). The ROC analysis of mRNA126 at UAC 0.976 had sensitivity of (0.80) and 1-specificity of (0.60) were total area under the curve = 759, pv=0.005; that means miRN-126 can be used as a screening test for CAD. Thus, miR126 can be considered as a biomarker on CAD in patient.

# 

\* Corresponding author: Hibah Rajaa Taher

⊠ E-mail: <u>heba.rajaie1105a@csw.uobaghdad.edu.iq</u>

© 2023 by SPC (Sami Publishing Company)

#### Introduction

Coronary Artery Disease (CAD) is a type of human cardiovascular disease resulted from thrombosis or acute coronary atherosclerosis leading to sudden cardiac death, CAD is the greatest popular form of cardiac disease [1]. Atherosclerosis (AS) is an inflammatory condition of the arteries marked by the development of atherosclerotic plaques causing CAD. These plaques cause the artery's lumen to constrict, which can eventually result in the production of thrombi and total obstruction of the lumen [2]. MicroRNA (miRNA) families are powerful regulators in CAD among other various diseases [3], as a main target of the current study having 18-22 nucleotides in single-stranded ncRNAs binding to the 3'-untranslated region of specific messenger RNA leading a decreasing in protein expression [4] because the particular miRNAs may be absorbed by the cells causing AS, these miRNAs can be considered as diagnostic biomarkers of AS [5] through whole blood, serum, or plasma. All related articles specified microRNAs linked to the particular molecular pathways that guide to AS, arterial remodeling, endothelial dysfunction, monocyte activation, arterial wall invasion, platelet activation, and vascular smooth muscle cell activation [6-8].

MiRNA-126 is abundant in endothelial cell may be treated as a biomarker for the CAD severity and complexity [9] with vascular integrity, angiogenesis, and wound healing. It exhibits several actions like atheroprotective, suppressing the inflammatory cascade, and mediating leukocyte adhesion in AS by reducing vascular cell adhesion expression [10, 11]. Low-density lipoprotein cholesterol (LDL-C) plays role in the progression and development of AS. Two forms of low-density lipoprotein cholesterol (LDL-C) controlled by miR-126 especially cholesterol metabolism of LDL-C [12] and regulated highdensity lipoprotein (HDL) metabolism and reverse cholesterol transport with direct targeting of cellular cholesterol efflux [13, 14]. IL-6 plays role in the development and maintenance of the inflammatory response in AS,

and

cytokine [15, 16]. CAD-related miRNAs (miR126)

immune-regulatory

pro-inflammatory,

are considered to be inflammatory-related miRNA influenced the inflammatory cytokines to coordinate the inflammatory processes [17]. Both cytokines and miRNA are elements of the signaling pathways that promote disease progression [18]. Also, higher sensitivity troponin (hs-Tn) levels were associated with multiple abnormalities of cardiac structure and function [19].

Therefore, this research aimed to study all of the above regulation elements and suggested them as good indictors of CAD in both genders of Iraqi patients compared with miR126.

#### **Materials and Methods**

#### Characteristics of patients

This study was carried out in Ibn Al-Bitar Hospital for Heart Surgery in Iraq from December 2020 to April 2021 for 8 patients of both sexes whose average age between 35-65 years old with CAD and 40 healthy people as a control group who have the same average age 35-65 year old. All patients were clinically diagnosed by the hospital doctor.

# Ethical considerations of this study

The volunteers were aware of the research, their names were not revealed to anyone, their findings were unharmed by their participation, and the research was done in accordance with our scientific committee.

# Anthropometric measurement

Body mass index (BMI) was determined by dividing the weight in kilograms by the height in square meters.

## Exclusion criteria

Patients who have already had liver cirrhosis, alcoholism, tobacco use, hypertension, hyperthyroidism, hypothyroidism, leukemia, cancer, obese, kidney disease, and COVID-19 were excluded from this study.

#### *Collection of blood samples*

Serum was separated from blood collected from venous blood samples (10 mL) were collected of

the studied subjects after fasting overnight. The serum is divided into three sections: a section for biochemistry measurements (lipid profile), ELISA (IL6), and high sensitive troponin (hs-Tn) measurements, respectively. Four Mls of the collected blood had been mixed with 0.5 mL of Trizole and kept under -70 °C to measure miRNA126.

#### Measurement of IL6

Enzyme Linked Immune Sorbent Assay (ELISA)/ sandwich technique were used for quantitative determination of Interleucken6, (IL-6). ELISA kits supplied by Shanghai Biological/China.

# Assay of high sensitive troponin

An automated quantitative test that uses the ELFA method to determine the presence of human cardiac troponin in human serum or plasma on devices from the VIDAS family (Enzyme Linked Fluorescent Assay) [20].

#### Biochemical examination

#### Determine lipid profile

Serum lipid profile (Total cholesterol (TC), serum triglycerides (TG), high density lipoprotein (HDL), low density lipoprotein (LDL), and very low density lipoprotein (VLDL)) determination occurs by standard assays were used in hospital by villas. (TC, TG, and HDL) are supplied by Abbott/ Germany. The lipid profile assays were performed by using a multi-channel Abbott Spectrum auto-analyzer (Abbott Laboratories, C460276, USA) according to the manufacturer instructions as opposed to LDL, which was determined by using the Friedewald Equation. LDL = (TC-HDL-[TG/5]), and VLDL was calculated by dividing total serum TG by 5 [21].

Isolation of total RNA and real-time polymerase chain reaction (RT-PCR) analysis

-SYBR green quantitative real-time polymerase chain reaction assay (RT-PCR) by using the miScript SYBR Green PCR kit, was applied to quantify miRNA-126 (Qiagen, Hilden, Germany). Mature miRNA sequence and a particular forward PCR primer were the primers' sequences.

(50CATTATTACTTTTGGTACGCG30) (50GGGGCATTATTACTTTTGG30).

The amplitude of miRNA expression change observed in patients in relation to control group was analyzed by the 22DDCt method [22].

RNA was isolated from the sample according to the protocol of TRIzol™ Reagent. For each tube, 0.4 mL of serum was added to 0.5 mL of TRIzol™ Reagent the lysate was homogenized. For each tube, 0.2 mL of chloroform was added to the lysate. All mixes were incubated, and then it was centrifuged for 10 minutes at 10.000 rpm. The mixture was separated into lower organic phase, interphase, and a colorless upper aqueous phase which containing the RNA.

Determination of RNA, cDNA yield fluorescence method [22]

QuantusFluorometer was used to detect the concentration of the extracted cDNA to detect the samples quality for downstream applications. For 1  $\mu$ l of cDNA, 199  $\mu$ l of diluted Quantifluor Dye was mixed. After 5 minute incubation at room temperature in dark place, cDNA concentration values were detected.

Gene expression by using pfaffi method [23] relative quantification:

Folding =  $2-\Delta\Delta CT$ 

 $\Delta$ CT = CT gene - CT House Keeping gene

 $\Delta\Delta CT = \Delta CT$  Treated or Control - Average  $\Delta CT$  Control.

## **Results and Discussion**

Mean±SD, and the (P-value) to the result for gender, age, BMI, hs-Tn, IL6, Chol, Tri, HDL, LDL, VLDL, and mir126 were listed in Table 1. Based on this table, it is obvious that there is a high significant differences between the (age, IL6, Chol, Tri, HDL, LDL, and VLDL) of control and patient groups and non-significant for (gander, BMI, hs-Tropon, miR126) at p  $\leq$  0.05 level, as presented in Table 1. The anthropometric and biochemical parameters of the paticipants are indicated in Table 1, where Age, IL6, Chol, Tri, HDL, LDL, and VLDL in research group showed statistically significant variations.

**Table 1:** Mean standard error (Mean ± SDE), standard deviation (Std. Deviation), P-value for the all criteria on patient and control groups under this study

	patient and control groups ander and stady							
		Mean±SDE	Std. Deviation	Sig. (P-value)				
Gander	patient	1.68±0.110	0.478	0.100				
	control	1.90±0.069	0.308					
Ago	patient	52.8947±2.14380	9.34460	0.007				
Age	control	44.5500±2.00588	8.97057	0.007				
BMI	patient	25.8363±0.42191 1.83906		0.600				
DIVII	control	25.4820±0.51444	2.30065	0.000				
hs-Tn	patient	2.2368±0.70551	3.07524	0.488				
(ng/L)	control	1.5790±0.62427	2.79182	0.400				
IL6	patient	166.88982±3.727898	15.370516	0.000				
(ng/L)	control	44.42247±4.704528	20.506564	0.000				
Chol	patient	172.5263±7.91415	34.49697	0.001				
(mg/dl)	control	129.7500±9.34813	37.39251					
Tri	patient	194.0588±29.23629	120.54432	0.001				
(mg/dl)	control	80.1765±5.85438	24.13824	0.001				
HDL	patient	39.5556±1.90153	8.06753	0.002				
(mg/dl)	control	53.5500±3.51648	15.72619					
LDL	patient	132.5000±9.49190	35.51544	0.002				
(mg/dl)	control	100.7222±3.65846	15.52154					
VLDL	patient	29.0000±3.29587	13.98318	0.002				
(mg/dl)	control	17.8000±1.44295	6.45307	0.003				
miR126	patient	0.6466316±0.23917814	1.04255334	0.240				
IIIIK120	control	1.1058438±0.30721843	1.22887374	0.240				

Control group of all these biochemical revealed a significant rise for results Age, IL6, Chol, Tri, HDL, LDL, and VLDL in the coronary artery (coronary heart) patients group that was consistent with Hedayatnia, M., *et al.* (2020) [24].

Heart disease patients' total serum Chol., Tri., LDL, and VLDL levels had been rising relative to the controls for all lipid profile indicators except HDL [25].

Likewise, the p-value for Il6 and Age showed an increase in the patient group compared with the healthy group. The pleiotropic cytokine IL6 has been found to be a prognostic indicator for many CVDs, including AF and CAD [26], while gender, BMI, hs-Tn, and miR126 indicated the apparent effect for both patients and other healthy people. Patients serum Tri has a positive correlation with BMI (r=0.567, pv=0.018), but it was negatively correlated with gender (r=0.606, pv=0.01). VLDL showed a positive correlation with Tri (r=0.860, pv=0.000) and a negative correlation with HDL (r=-0.741, pv=0.001) (Table 2). In control group, Pearson correlation showed:

\*A positive correlation between Chol and Age (r=0.515, pv=0.41), and between VLDL and Tri. \*Conversely, the negative correlation shows between Tri and hs-Tn (r=-0.521, pv=0.032), Tri and IL6 (r=-0.754, pv=0.001), and between VLDL and IL6 (r=-0.494, pv=0.031).

Based on Tables 2 and 3 (The pearson correlation of patients and control groups) demonstrated miRNA-126 and lipid characteristics did not significantly correlate in any studies subject that were agreed with Ortega *et al.* [27] and they were disagreed with G. Musso *et al.* [28].

#### ROC curve and cute-off analysis

Receiver Operating Characteristic (ROC) curve and the best cut-off point for mRN126 at 0.976 had a sensitivity of (0.80) and 1-specificity of (0.60) were the total area under the curve = 759, pv=0.005. It means that miR126 at this cut-off point can be used as a screening test to detect patients with coronary artery diseases, as displayed in Figure 1 and Table 4.

**Table 2:** Pearson correlation for two sets of data for patients population

	Correlations (patient group)											
		Gander	Age	BMI	h.sTropon	IL6	Chol	Tri	HDL	LDL	VLDL	miR12 6
Gander	r	1	-0.207	-0.396	-0.416	0.059	0.014	-0.606**	0.276	-0.402	-0.256	-0.377
Galluel	p		0.395	0.093	0.076	0.823	0.955	0.010	0.268	0.154	0.306	0.112
Age	r		1	0.172	0.134	0.183	-0.121	0.035	-0.001	0.319	0.088	-0.378
Age	p			0.481	0.586	0.482	0.622	0.893	0.998	0.266	0.728	0.111
BMI	r			1	0.299	-0.260	0.196	0.567*	-0.169	0.122	0.164	0.076
DIVII	p				0.214	0.314	0.421	0.018	0.503	0.678	0.515	0.756
hs-Tn	r				1	-0.288	0.105	0.254	-0.224	0.028	0.299	-0.080
115-111	p					0.263	0.668	0.326	0.371	0.925	0.228	0.745
IL6	r					1	-0.057	-0.243	0.147	0.200	-0.076	0.324
	p						0.829	0.384	0.586	0.533	0.780	0.205
Chol	r						1	0.327	-0.177	0.103	0.168	-0.341
CHOI	p							0.200	0.483	0.727	0.504	0.153
Tri	r							1	-0.385	0.388	0.860**	0.113
111	p								0.141	0.212	0.000	0.666
HDL	r								1	-0.423	-0.741**	-0.002
прь	p									0.149	0.001	0.995
LDL	r									1	0.203	0.372
гъг	p		_	_	_	_				_	0.486	0.191
VLDL	r										1	-0.284
4 LDL	p											0.253
miR126	r											1
111IK120	p											

<sup>\*\*</sup> Correlation is significant at the 0.01 level (2-tailed)

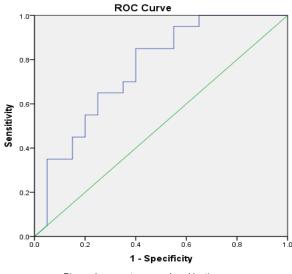
Table 3: Pearson correlation of control set

	Correlations (control)											
		Gander	Age	BMI	h.sTropon	IL6	Chol	Tri	HDL	LDL	VLDL	miR126
Gander	r	1	-0.208	0.411	0.191	-0.111	0.241	-0.075	0.208	-0.294	0.042	0.188
danuei	p		0.379	0.072	0.420	0.651	0.369	0.774	0.380	0.237	0.859	0.485
Age	r		1	0.152	0.140	-0.129	0.515*	0.209	0.086	0.009	0.265	0.190
Age	p			0.523	0.556	0.600	0.041	0.421	0.719	0.973	0.259	0.480
BMI	r			1	0.277	-0.217	0.362	0.126	0.189	0.085	0.106	0.400
DIVII	p				0.238	0.371	0.168	0.631	0.424	0.739	0.657	0.125
hs-Tn	r				1	0.096	0.341	-0.521*	0.095	-0.232	-0.074	0.244
115-111	p					0.695	0.196	0.032	0.691	0.354	0.755	0.362
IL6	r					1	0.242	-0.754**	0.282	0.043	-0.494*	-0.135
	p						0.385	0.001	0.242	0.871	0.031	0.619
Chol	r						1	-0.326	-0.144	-0.185	-0.028	-0.005
	p							0.256	0.595	0.492	0.917	0.987
Tri	r							1	-0.156	0.247	0.946**	0.114
	p								0.551	0.356	0.000	0.711
HDL	r								1	-0.026	-0.074	0.182
	p									0.917	0.758	0.499
LDL	r									1	-0.281	0.256
	p										0.259	0.378
VLDL	r										1	0.101
	p		-			-		-				0.711
miR126	r											1
mintizo	p											

<sup>\*</sup> Correlation is significant at the 0.05 level (2-tailed)

<sup>\*</sup> Correlation is significant at the 0.05 level (2-tailed)

<sup>\*\*</sup> Correlation is significant at the 0.01 level (2-tailed)



Diagonal segments are produced by ties

Figure 1: ROC curve

Table 4: The use of miR126 for detecting patients with coronary artery diseases

	Area Under the Curve Test Result Variable(s): miR126		_	ptotic95% ence Interval	miR 126 cutoff value	Sensitivity	Specificity	
	Area	Std.Error	Asymptotic sig.	Low Bound	Upper Bound	0.976	80%	60%
ĺ	0.759	0.077	0.005	0.609	0.909			

All the samples under this study (patients and healthy people) had the body mass less than 25. Therefore, the obesity and high weight were excluded, which had led body mass was not affected factor. Often associated with CAD are comorbidities such as abnormal lipid profiles which then leads to the dyslipidemia development and the well-known lipid triad: high levels of plasma triglycerides, low levels of highdensity lipoprotein, and the appearance of small dense low-density lipoproteins (dsLDL). This triad is along with endothelial dysfunction, which can also be induced and contributed to atherosclerotic plaque formation. Likewise, lipid triad inhibits lipoprotein lipase activity in adipocytes leading to an increased release of free fatty acids and inflammatory cytokines such as IL-6. This explains the reason for the rise of lipid (Tri, Chol, VLDL, and dsLDL) and IL-6, and decrease of HDL in atherosclerosis [29]. Age showed an increase in the patient group (patient with CAD) in comparison with the healthy group;

Since age is a risk factor for getting CAD, and includes age >45 years old in men and >55 years old in women [30]. Men are more likely to develop coronary heart disease (CHD) than women that they tend to do at a younger age than women. This is closely related to the estrogen deficiency, due to ovarian failure, endothelial dysfunction, where estrogen acts as an antiarteriosclerosis in women in their early to midlife. However, in this study, there was no significant difference between the gender in both patient group and control group. Because younger women accounted for 80% of all the healthy and patients [31, 32]. After myocardial damage begins, cardiac troponin I (cTnI) levels rise quickly and return to normal within 7 days [33]. Acute myocardial infarction (AMI), cardiac arrhythmias, and heart failure are acute and chronic cardiac disease states associated with elevated cTn concentrations. cTn is a reliable of cardiovascular and mortality in the general population3-5, in people

with known CAD, acute coronary syndrome (ACS), non-cardiac disease, and in people without these conditions [34]. The cases under this study all had CHD, but they had a history on CHD for a long time. Thus, there was no effect on troponin tests, which proves that troponin levels rise in blood when the heart attack is acute and immediately [35]. The miR-126 level did not change so no significant correlation was found between control and patient groups (p = 0.274). This might be brought on by the study participants' regular use of aspirin and -blockers, which could have an impact on the miR-126 level [36]. These results suggest that miRNA levels were unaffected by anti-platelet treatments [37, 38] that was agreed with Fiedler J (2011) [39] and disagreed with Mishra S [35]. It is possible to classify LDL cholesterol as a typical proatherogenic substance. Normally, LDLreceptors remove LDL particles from circulation (LDL-R). Different lipid metabolism-related molecular pathways are impacted hyperlipidemia CAD. The LDL particles circulate in the bloodstream for an extended period of time [40, 41]. The reverse cholesterol transport or the cholesterol movement from peripheral tissues like macrophages in atherosclerotic plaques back to the liver is made by HDL particles. Thus, patients with hyperlipidemia and CAD may benefit from the enhancement of reverse cholesterol transport via HDL [41]. MiRNAs have been demonstrated to have important regulatory functions in lipid metabolism. They play a key role in coordinating the normal progression of the lipid metabolism. Since the use of apoptotic bodies caused the atherosclerotic plaques to regress, the miR-126 application also appears to be highly promising. Nevertheless, this is due to the possibility that it could have a proatherogenic effect by activating vascular smooth muscle cells [42]. In the present study, there is no significant correlation between plasma levels of miRNA-126 in diseased groups and the LDL levels and other lipid; uncertainty persists regarding mechanism underlying the proportional drop in miR126 levels in the blood among CAD patients and the rise in LDL cholesterol. Despite the fact that high-density lipoprotein (HDL) cholesterol

and miRNA are known to be connected. The transcription, processing, and stability of the miRNAs within circulating cells, as well as the capacity of these cells to release miR NAs into the plasma, may all have an impact on the amounts of circulating miRNAs. Circulating miRNAs may be delivered to the heart or blood vessel cells via microvesicles, exosomes, or apoptotic bodies. Since the current study evaluated miR-126, the current findings in plasma could be a result of modifications in miRNA transcription, processing, and their release from circulating cells. It is yet unknown if down-regulation of miR-126 in CAD patients is a direct cause of inflammation or a response to it. Based on the observed variations in miR-126 level in relation to LDL cholesterol, it is possible that the levels of circulating miRNA are a result of a compensatory response to inflammation in the hyperlipidemia context. Endothelial miR-126 may become depleted as vascular disease processes progress and plasma levels may begin to drop. Transcoronary transit consumes the vascular miR-126. According to certain reports, specific miRNAs control cholesterol efflux. However, it is not anticipated that miR-126 will target any LDL component(s). Statin did not affect miR-126 in endothelial progenitor cells, despite the fact that statin was often administered to CAD patients in this study. The underlying processes that may connect LDL cholesterol and circulating miR-126 levels in patients with CAD need to be further investigated [43].

#### **Conclusion**

Micro RNAs as important regulators of the course of cardiac disease serve as the early indicators and therapeutic targets for cardiometabolic disease. Elevation of some miRNAs such as miR126 in cardiovascular patients can efficiently prognoses the CAD development in these patients. Therefore, miRNA126 can be used as a biomarker because its level was significantly decreased in patients with CAD.

#### **Acknowledgments**

My thanks and gratitude to all those who helped me carry out this research.

#### **Funding**

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

#### **Authors' contributions**

All authors contributed to data analysis, drafting, and revising of the paper and agreed to be responsible for all the aspects of this work.

#### **Conflict of Interest**

There are no conflicts of interest in this study.

#### References

- [1]. Asal G.M.H., Al-Janabib L.M., Hussein N.Y., Study of the participation of Lipoprotein Lipase gene polymorphism in coronary artery disease; *Eurasian Chemical Communications*, 2022, **4**:590 [Crossref], [Google Scholar], [Publisher]
- [2]. Mahjoob G., Ahmadi Y., Fatima rajani H., Khanbabaei N., Abolhasani S., Circulating microRNAs as predictive biomarkers of coronary artery diseases in type 2 diabetes patients, *Journal of Clinical Laboratory Analysis*, 2022, 36:e24380 [Crossref], [Google Scholar], [Publisher]
- [3]. Saliminejad K., Khorram Khorshid H.R., Soleymani Fard S., Ghaffari S.H., An overview of microRNAs: biology, functions, therapeutics, and analysis methods, *Journal of Cellular Physiology*, 2019, 234:5451 [Crossref], [Google Scholar], [Publisher]
- [4]. Beilerli A., Gareev I., Beylerli O., Yang G., Pavlov V., Aliev G., Ahmad A., Circular RNAs as biomarkers and therapeutic targets in cancer, *Seminars in Cancer Biology*, 2022, **83**:242 [Crossref], [Google Scholar], [Publisher]
- [5]. Libby P., Ridker P.M., Hansson G.K., Progress and challenges in translating the biology of atherosclerosis, *Nature*, 2011, **473**:317 [Crossref]
  [6]. Taurino C., Miller W.H., McBride M.W., McClure J.D., Khanin R., Moreno M.U., Dymott J.A., Delles C., Dominiczak A.F., Gene expression profiling in whole blood of patients with coronary artery disease, *Clinical Science*, 2010, **119**:335 [Crossref], [Google Scholar], [Publisher]

[7]. Siasos G., Bletsa E., Stampouloglou P.K., Oikonomou E., Tsigkou V., Paschou S.A., Vlasis K., Marinos G., Vavuranakis M., Stefanadis C., Tousoulis D., MicroRNAs in cardiovascular disease, *Hellenic Journal of Cardiology*, 2020, 61:165 [Crossref], [Google Scholar], [Publisher] [8]. The role of inflammation in cardiovascular diseases: the predictive value of neutrophillymphocyte ratio as a marker in peripheral arterial disease, *Therapeutics and Clinical Risk* 

Management, 2016, 12:851 [Crossref], [Google

Scholar], [Publisher]

- [9]. Sheikh M.S.A., Almaeen A., Alduraywish A., Alomair B.M., Salma U., Fei L., Yang T.L., Overexpression of miR-126 Protects Hypoxic-Reoxygenation-Exposed HUVEC Cellular Injury through Regulating LRP6 Expression, *Oxidative Medicine and Cellular Longevity*, 2022, 2022:3647744 [Crossref], [Google Scholar], [Publisher]
- [10]. Solly E.L., Dimasi C.G., Bursill C.A., Psaltis P.J., Tan J.T., MicroRNAs as therapeutic targets and clinical biomarkers in atherosclerosis, *Journal of Clinical Medicine*, 2019, **8**:2199 [Crossref], [Google Scholar], [Publisher]
- [11]. Harris T.A., Yamakuchi M., Kondo M., Oettgen P., Lowenstein C.J., Ets-1 and Ets-2 Regulate the Expression of MicroRNA-126 in Endothelial Cells, *Arteriosclerosis, Thrombosis, and Vascular Biology*, 2010, **30**:1990 [Crossref], [Google Scholar], [Publisher]
- [12]. Jin X., Yang S., Lu J., Wu M., Small, dense low-density lipoprotein-cholesterol and atherosclerosis: relationship and therapeutic strategies, *Frontiers in Cardiovascular Medicine*, 2021, **8**:804214 [Crossref], [Google Scholar], [Publisher]
- [13]. Khan A.A., Gupta V., Mahapatra N.R., Key regulatory miRNAs in lipid homeostasis: Implications for cardiometabolic diseases and development of novel therapeutics, *Drug Discovery Today*, 2022, **27**:2170 [Crossref], [Google Scholar], [Publisher]
- [14]. Esau C., Davis S., Murray S.F., Yu X.X., Pandey S.K., Pear M., Watts L., Booten S.L., Graham M., McKay R., Subramaniam A., Propp S., Lollo B.A., Freier S., Bennett C.F., Bhanot S., Monia B.P., miR-122 regulation of lipid metabolism

revealed by in vivo antisense targeting, *Cell Metabolism*, 2006, **3**:87 [Crossref], [Google Scholar], [Publisher]

[15]. Ridker P.M., From C-reactive protein to interleukin-6 to interleukin-1: moving upstream to identify novel targets for atheroprotection, *Circulation Research*, 2016, **118**:145 [Crossref], [Google Scholar], [Publisher]

Lu S., Wang Y., Wang Y., Hu J., Di W., Liu S., Zeng X., Yu G., Wang Y., Wang Z., The IL-6 rs1800795 and rs1800796 polymorphisms are associated with coronary artery disease risk, Journal of Cellular and Molecular Medicine, 2022, **24**:6191 [Crossref], [Google Scholar], [Publisher] [17].Gorabi A.M., Kiaie N., Sathyapalan T., Al-Rasadi K., Jamialahmadi T., Sahebkar A., The Role of MicroRNAs in Regulating Cytokines and Growth Factors in Coronary Artery Disease: The Ins and Outs, Journal of Immunology Research, 2020, **2020**:5193036 [Crossref], [Google Scholar], [Publisher]

Sadri Moghoofei [18]. Nahand J., M., Salmaninejad A., Bahmanpour Z., Karimzadeh M., Nasiri M., Mirzaei H.R., Pourhanifeh M.H., Bokharaei-Salim F., Mirzaei H., Hamblin M.R., Pathogenic role of exosomes and microRNAs in HPV-mediated inflammation and cervical cancer: a review, International Journal of Cancer, 2020, **146**:305 [Crossref], [Google Scholar], [Publisher] Sherwood M.W., Kristin [19]. Newby L., High-sensitivity troponin assays: evidence, indications, and reasonable use, Journal of the American Heart Association, 2014, 3:e000403 [Crossref], [Google Scholar], [Publisher]

[20]. Somlor S., Brossault L., Grandadam M., Evaluation of VIDAS® Diagnostic Assay Prototypes Detecting Dengue Virus NS1 Antigen and Anti-Dengue Virus IgM and IgG Antibodies, *Diagnostics*, 2021, **11**:1228 [Crossref], [Google Scholar], [Publisher]

[21]. Maki K.C., Grant J.K., Orringer C.E., LDL-C Estimation: The Perils of Living With Imperfection, *Journal of the American College of Cardiology*, 2022, **79**:542 [Crossref], [Google Scholar], [Publisher]

[22]. Barseem N.F., Mahasab M.M., Zaed I.F., Said A.E.A., Abd El Gayed E.M., Genetic Indices Relationship to Hyperglycemia-associated

Biomarkers: Consistency with miRNA Expression in Egyptian Children with T1DM, Journal of Clinical Research in Pediatric Endocrinology, 2022, **14**:PP76 [Crossref], [Google Scholar], [Publisher] [23]. Ng H.F., Ngeow Y.F., Α simple spreadsheet-based method for relative quantification using quantitative real-time PCR, Biochemistry and Molecular Biology Education, **50**:99 [Crossref], [Google Scholar], 2022, [Publisher]

[24]. Hedayatnia M., Asadi Z., Zare-Feyzabadi R., Yaghooti-Khorasani M., Ghazizadeh H., Ghaffarian-Zirak R., Nosrati-Tirkani A., Mohammadi-Bajgiran M., Rohban M., Sadabadi F., Rahimi H.R., Ghalandari M., Ghaffari M.S., Yousefi A., Pouresmaeili E., Besharatlou M.R., Moohebati M., Ferns G.A., Esmaily H., Ghayour-Mobarhan M., Dyslipidemia and cardiovascular disease risk among the MASHAD study population, *Lipids Health Dis*, 2020, **19**:42 [Crossref], [Google Scholar], [Publisher]

Kim H.Y., Hong M.H., Kim K.W., Yoon J.J., [25]. Lee J.E., Improvement of Hypertriglyceridemia by Roasted Nelumbinis folium in High Fat/High Cholesterol Diet Rat Model, Nutrients, 2020, **12**:3859 [Crossref], [Google Scholar], [Publisher]. Rosa M., Chignon A., Li Z., Boulanger C., [26]. Arsenault B.J., Bossé Y., Thériault S., Mathieu P., A Mendelian randomization study of IL6 signaling in cardiovascular diseases, immune-related disorders and longevity, npj Genomic Medicine, 2019, **4**:23 [Crossref], [Google Scholar], [Publisher]

[27]. Ortega F.J., Mercader J.M., Moreno-Navarrete J.M., Rovira O., Guerra E., Esteve E., Xifra G., Martínez C., Ricart W., Rieusset J., Rome S., Karczewska-Kupczewska M., Straczkowski M., Fernández-Real J.M., Profiling of circulating microRNAs reveals common microRNAs linked to type 2 diabetes that change with insulin sensitization, *Diabetes Care*, 2014, **37**:1375 [Crossref], [Google Scholar], [Publisher].

[28]. Musso G., Gambino R., Cassader M., Cholesterol metabolism and the pathogenesis of non-alcoholic steatohepatitis, *Progress in Lipid Research*, 2013, **52**:175 [Crossref], [Google Scholar], [Publisher]

- [29]. Ormazabal V., Nair S., Elfeky O., Aguayo C., Salomon C., Zuñiga F.A., Association between insulin resistance and the development of cardiovascular disease, *Cardiovascular Diabetology*, 2018, **17**:122 [Crossref], [Google Scholar], [Publisher]
- [30]. Gao Z., Chen Z., Sun A., Deng X., Gender differences in cardiovascular disease, *Medicine in Novel Technology and Devices*, 2019, **4**:100025 [Crossref], [Google Scholar], [Publisher]
- [31]. Hajar R., Risk Factors for Coronary Artery Disease: Historical Perspectives, *Heart views: the official journal of the Gulf Heart Association*, 2017, **18**:109 [Crossref], [Google Scholar], [Publisher]
- [32]. Zhu D., Chung H.F., Dobson J.A., Pandeya N., Giles G.G., Age at natural menopause and risk of incident cardiovascular disease: a pooled analysis of individual patient data, *The Lancet Public Health*, 2019, **4**:e553 [Crossref], [Google Scholar], [Publisher]
- [33]. Tiwari R.P., Jain A., Khan Z., Kohli V., Bharmal R.N., Kartikeyan S., Bisen P.S., Cardiac Troponins I and T: Molecular Markers for Early Diagnosis, Prognosis, and Accurate Triaging of Patients with Acute Myocardial Infarction, *Molecular Diagnosis & Therapy*, 2012, **16**:371 [Crossref], [Google Scholar], [Publisher]
- [34]. Tveit S.H., Myhre P.L., Hanssen T.A., Forsdahl S.H., Iqbal A., Omland T., Schirmer H., Cardiac troponin I and T for ruling out coronary artery disease in suspected chronic coronary syndrome, *Scientific Reports*, 2022, **12**:945 [Crossref], [Google Scholar], [Publisher]
- [35]. Lippi G., Cervellina G., Sanchis-Gomara F., Predicting mortality with cardiac troponins: recent insights from meta-analyses, 2019 [Crossref], [Google Scholar], [Publisher]
- [36]. Mishra S., Rizvi A., Pradhan A., Perrone M.A., Ali W., Circulating microRNA-126 &122 in patients with coronary artery disease: Correlation with small dense LDL, *Prostaglandins & Other Lipid Mediators*, 2021, **153**:106536 [Crossref], [Google Scholar], [Publisher]

- [37]. Wang X., Lian Y., Wen X., Guo J., Wang Z., Jiang S., Hu Y., Expression of miR-126 and its potential function in coronary artery disease, *African Health Sciences*, 2017, **17**:474 [Crossref], [Google Scholar], [Publisher]
- [38]. Willeit P., Zampetaki A., Dudek K., Kaudewitz D., King A., Kirkby N.S., Crosby-Nwaobi R., Prokopi M., Drozdov I., Langley S.R., Sivaprasad S., Markus H.S., Mitchell J.A., Warner T.D., Kiechl S., Mayr M., Circulating microRNAs as novel biomarkers for platelet activation, *Circulation Research*, 2013, **112**:595 [Crossref], [Google Scholar], [Publisher]
- [39]. Fiedler J., Jazbutyte V., Kirchmaier B.C., Gupta S.K., Lorenzen J., Hartmann D., et al. MicroRNA-24 regulates vascularity after myocardial infarction, *Circulation*, 2011, 9124:720 [Crossref], [Google Scholar], [Publisher]
- [40]. Ginter E., Simko V., New promising potential in fighting atherosclerosis: HDL and reverse cholesterol transport, *Bratislavské lekárske Listy*, 2013, **114**:172 [Crossref], [Google Scholar], [Publisher]
- [41]. Peluso I., Morabito G., Urban L., Ioannone F., Serafi M., Oxidative stress in atherosclerosis development: the central role of LDL and oxidative burst, *Endocrine, Metabolic and Immune Disorders Drug Targets*, 2012, **12**:351 [Crossref], [Google Scholar], [Publisher]
- [42]. Zernecke A., Bidzhekov K., Noels H., Shagdarsuren E., Gan L., Denecke B., Hristov M., Köppel T., Jahantigh M.N., Lutgens E., Wang S., Olson E.N., Schober A., Weber C., Delivery ofmicroRNA-126 by apoptotic bodies induces CXCL12-dependent vascular protection, *Science Signaling*, 2009, **2**:ra81 [Crossref], [Google Scholar], [Publisher]
- [1]. Sun X., Zhang M., Sanagawa A., Mori C., Ito S., Iwaki S., Satoh H., Fujii S., Circulating microRNA-126 in patients with coronary artery disease: correlation with LDL cholesterol, *Thrombosis Journal*, 2012, **10**:16 [Crossref], [Google Scholar], [Publisher]

#### **HOW TO CITE THIS ARTICLE**

Hibah Rajaa Taher, Perry Habib Saifullah. Role of MiRNA126 As a Biomarker for Coronary Artery Disease in Iraqi Patients. *J. Med. Chem. Sci.*, 2023, 6(5) 1163-1172

https://doi.org/10.26655/JMCHEMSCI.2023.5.22

URL: http://www.jmchemsci.com/article\_160027.html