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## **Original Research Article**

# A gallic acid food electrochemical sensor based on amplification of paste electrode by Cdo/CNTs nanocomposite and ionic liquid

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### ABSTRACT

Gallic acid is one of the most abundant phytochemical in nature with anticancer activity against the prostate cancer. In this research work, carbon paste electrode (PE) modified with CdO/CNTs nanocomposite and 1-hexyl-3-methyl imidazoliume bromide (HMIZBr) design and made-up for determination of gallic acid in food samples. Electrochemical behavior of gallic acid at the CdO/CNTs/HMIZBr/PE was investigated in aqueous solution using the voltammetric methods. The gallic acid oxidation signal was improved about 2.82 times on the surface of the CdO/CNTs/HMIZBr/PE compared with that of the PE. Using differential pulse voltammetric method as sensitive strategy, the CdO/CNTs/HMIZBr/PE showed linear dynamic range 0.004-500 µM with detection limit of 0.9 nM to determine the gallic acid. In addition, real sample analysis data showed the powerful ability of the CdO/CNTs/HMIZBr/PE to determine the gallic acid in white rice.

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#### **Graphical Abstract**



#### Introduction

Gallic acid with antioxidant activity showed great effect against the cancer cells, which confirmed importance of the gallic acid in human body [1]. Gallic acid with phenolic structure is a trihydroxybenzoic acid that is found in sumac, witch hazel, gallnuts, tealeaves, and oak bark many types of plants [2, 3]. This compound has attracted a great deal of attention due to its application as antioxidant [4-10]. Gallic acid is useful in food industries such as the chocolate and the wine industry [11, 12]. Due to the aforementioned points, many analytical sensors were suggested to determine the gallic acid in food and pharmaceutical formulation [13-15].

Among the analytical methods, electrochemical methods revealed more advantages for determination of food and pharmaceutical compounds due to its fast response [16-18]. On the other hand, for easy determination of food and pharmaceutical compounds, researchers need to portable systems such as electrochemical sensors [20-

23]. To improve the electrochemical sensor ability, researchers modified them by conductive mediators such as ionic liquids and nanomaterials. Nanotechnology is a new approach to science and especially analytical sensors [24-26]. Due to high surface area of nanomaterials, they showed good advantages to modify the electrochemical sensors for electroactive compounds analysis [27]. Ionic liquids are another class of high-conductivity modifiers that have been widely used in recent years to modify the electrochemical sensors [28-32]. They are a suitable alternative binder for paraffin oil in fabrication of carbon paste modified electrodes.

In this research, a two-fold amplified sensor was designed and fabricated (CdO/CNTs/HMIZBr/PE) to determine the gallic acid in food samples. The oxidation signal of gallic acid was selected as analytical issue to determine antioxidant in food samples. Results showed that the CdO/CNTs/HMIZBr/PE is a good analytical sensor to determine the gallic acid in food samples.

#### Experimental

#### Reagents and Apparatus

The gallic acid (97%), 1-hexyl-3-methyl imidazoliume bromide, phosphoric acid, graphite powder, SWCNTs-COOH, sodium hydroxide, cadmium (II) acetate anhydrous and paraffin oil were purchased from the Merck and Sigma-Aldrich Company. CdO/CNTs were synthesis by reported procedure [33]. A µ-Autolab system (Netherland with NOVA software) was used to record the electrochemical signals. Ag/AgCl/KCl<sub>sat</sub> was used as reference electrode.

#### Preparation of CdO/CNTs/HMIZBr/PE

The CdO/CNTs/HMIZBr/PE was fabricated by mixing 0.06 g CdO/CNTs nanocomposite+0.94 g graphite powder and using paraffin oil+HMIZBr as binders with ration 80:20 v:v. The mixture was converted to a paste using hand mixing.

#### **Real Samples Preparation**

White rice samples were used to study the ability of the sensor in real sample analysis. 4.0 g of rice was powdered and then ultrasonic in 50% ethanol solution for 2 h. After filtering the sample, it was used for analysis of gallic acid using standard addition method.

#### **Result and Discussion**

#### Voltammetric Examination

Electro-oxidation of the gallic acid was evaluated at different pH of phosphate buffer solution (PBS) using CdO/CNTs/HMIZBr/PE. Cyclic voltammograms of 300  $\mu$ M gallic acid in the pH range 50-8.0 are presence in Figure 1 inset. A negative shift with slope 63.3 mV/pH (plot of E-pH) was observed for electrooxidation of the gallic acid, confirming the equal value of electron and proton in redox mechanism of this antioxidant compound (Figure 1).



**Figure 1.** Plot of potential-pH for electro-oxidation of 300 µM gallic acid (n=4). Inset) relative cyclic voltammograms.

On the other hand and using comparing cyclic voltammograms currents (Figure 2), maximum oxidation current for electro-oxidation of 300  $\mu$ M gallic acid using CdO/CNTs/HMIZBr/PE as

sensor was observed at pH=7.0 and this condition was used for the next steps of investigation.



To evaluate the type movement of gallic acid of this reaction, linear sweep voltammograms (LSV) of 250  $\mu$ M gallic acid was recorded at the scan rate ranging from 10 to 100 mV/s using the CdO/CNTs/HMIZBr/PE as electrochemical sensor (Figure 3). In continuous, the plot of current vs. v<sup>1/2</sup> was draw (Figure 3) and results confirmed the linear relation with equation I=2.8189 v<sup>1/2</sup>+3.2131 (R<sup>2</sup>=0.9933) for this investigation. This result confirmed a diffusion process for electro-oxidation of gallic acid on the surface of the CdO/CNTs/HMIZBr/PE. In addition, LSV of 400  $\mu$ M gallic acid was recorded on the surface of CPE (curve a), CdO/CNTs/PE (curve b), HMIZBr/PE (curve c) and CdO/CNTs/HMIZBr/PE (curve d). By moving from CPE to CdO/CNTs/HMIZBr/PE oxidation current of gallic acid was improved from 16.86  $\mu$ A to 47.6  $\mu$ A, confirming the positive role of the CdO/CNTs and HMIZBr in modification of CPE.



250 µM gallic acid at scan rates a) 10; b) 15; c) 30; d) 60 and e) 100 mV/s.



The linear dynamic range and limit of detection for determination of gallic acid using CdO/CNTs/HMIZBr/PE was recorded in this step. So, differential pulse voltammetric method was used (Figure 5). The signals confirmed a

linear dynamic range  $0.004-500 \mu$ M with detection limit of 0.9 nM for determining the gallic acid using CdO/CNTs/HMIZBr/PE (Figure 5).



CdO/CNTs/HMIZBr/PE (n=4). Inset) DP voltammograms of a) 0.004; b) 0.1; c) 10; d) 30; e) 50; f) 100; g) 150; h) 180; i) 250; j) 320; k) 400 and l) 500 µM gallic acid.

After identifying and optimizing the conditions of analysis, the ability of CdO/CNTs/HMIZBr/PE determine the gallic acid in real sample was checked. Obtained data

are demonstrated in Table 1. The recovery data confirmed the ability of the CdO/CNTs/HMIZBr/PE determine the gallic acid in real sample.

Table 1. Determination of gallic acid in food samples (n=3).				
samples	Added gallic acid	Expected gallic acid	Founded gallic acid	Recovery %
	(µM)	(µM)	(µM)	
White rice			<lod< td=""><td></td></lod<>	
	10.00	10.00	10.32±0.48	103.2
	20.00	20.00	19.78±0.56	98.9

#### Conclusion

An electrochemical strategy was selected as analytical approach to determine the gallic acid. The CdO/CNTs/HMIZBr/PE was selected as working electrode in an electrochemical system. The CdO/CNTs/HMIZBr/PE showed high performance ability for determination of gallic acid with detection limit 0.9 nM. In addition, CdO/CNTs/HMIZBr/PE was used to determine the gallic acid in white rice as real sample. Real sample analysis data showed a recovery range 98.9-102.2% that is sufficient for a new and high quality food sensor.

#### References

[1] Calheiros R., Fiuza S., Gomes C., Marques M., Milhazes N., Borges F. *Fund. Clin. Pharmacol.*, 2004, **18**:120

[2] Fiuza S., Gomes C., Teixeira L., Da Cruz M.G., Cordeiro M., Milhazes N., *Bioorg. Med. Chem.*, 2004, **12**:3581

[3] Goulas V., Stylos E., Chatziathanasiadou M.V., Mavromoustakos T., Tzakos A.G. *Int. J. Mol. Sci.*, 2016, **17**:1875

[4] Pathak S., Niranjan K., Padh H., Rajani M. *Chromatographia.*, 2004, **60**:241

[5] Koyama K., Goto-Yamamoto N., Hashizume K. Biosci. Biotech. Bioch., 2007, 71:958 [6] Pandurangan A.K., Mohebali N., Norhaizan M.E., Looi C.Y. Drug Des. Dev. Ther., 2015, 9:3923 [9] Alemika T.E., Onawunmi G.O., Olugbade T. Niger. J. Nat. Prod. Med., 2006, 10:108 [10] Chanwitheesuk A., Teerawutgulrag A., Kilburn J.D., Rakariyatham N. Food Chem., 2007, **100:**1044 [11] Nakai S., Inoue Y., Hosomi M., Murakami A. Water Res., 2000, 34:3026 [12] Zucca P., Rosa A., Tuberoso C.I., Piras A., Rinaldi A.C., Sanjust E., Nutrients., 2013, 5:149 [13] Shahrzad S., Bitsch I. J. Chromatogr. B., 1998, **705**:87 [14] Abdel-Hamid R., Newair E.F. J. Electroanal. Chem., 2013, 704: 32 [15] Vijayalakshmi R., Ravindhran R. Asian Pac. *J. Trop. Biomed.*, 2012, **2:**S1367 [16] Arabali V., Malekmohammadi S., Karimi F. Microchem. J., 2020, 158: 105179 [17] Baghayeri M., Rouhi M., Lakouraj M.M., Amiri-Aref M., J. Electroanal. Chem., 2017, 784: 69 [18] Baghayeri M., Alinezhad H., Fayazi M., Tarahomi M., Ghanei-Motlagh R., Maleki B., Electrochim. Acta., 2019, 312: 80 [19] Maleki B., Baghayeri M., Abadi S.A.J., Tayebee R., Khojastehnezhad A., RSC Adv., 2016, **6:**96644 [20] Golikand A.N., Raoof J., Baghayeri M., Asgari M., Irannejad L. Russ. J. Electrochem., 209, **45:**192 [21] Baghayeri M., Ansari R., Nodehi M., Razavipanah I., Veisi H., Microchim. Acta, 2018, 185:320

[22] Hojjati-Najafabad A., Rahmanpour M.S., Karimi F., Zabihi-Feyzaba H., Malekmohammadi S., Agarwal S., Gupta V.K., *Int. J. Electrochem. Sci.*, 2020, **15**: 6969

[23] Zabihpour T., Shahidi S.A., Karimi Maleh H., Ghorbani-HasanSaraei A. *Eurasian Chem. Commun.*, 2020, **2**:362

[24] Baghayeri M., Mahdavi B., Hosseinpor-Mohsen Abadi Z., Farhadi S., *Appl. Organomet. Chem.*, 2018, **32**: e4057

[25] Targhoo A., Amiri A., Baghayeri M., *Microchim Acta.*, 2018, **185**:15

[26] Alizadeh M., Azar P.A., Mozaffari S.A.,Karimi-maleh H., Tamaddon A.M. *Front. Chem.*,2020, 8: 677

[27] Davarnia B., Shahidi S.A., Karimi-Maleh H., Ghorbani-HasanSaraei A., Karimi F., *Int. J. Electrochem. Sci.*, 2020, **15**:2549

[28] Fouladgar M. *J. Electrochem. Soc.,* 2018, **165** (13): B559

[29] Fouladgar M. *Food Anal. Methods.*, 2017, **10**:1507

[30] Fouladgar M. *Sens. Actuators B Chem.*, 2016, **230:** 456

[31] Baghayeri M., Sedrpoushan A., Mohammadi A., Heidari M. *Ionics.,* 2017, **23**: 1553

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