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Role of Lymphocytes and Atyphical Lymphocytes in Dengue Hemorrhagic Fever: A Literature Review

Ahmad Fahrudi Setiawan^{1*} , Yuyun Yueniwati Prabowowati Wajib², Kusworini Handono³, Setyawan Purnomo Sakti⁴

¹Doctoral Program of Medical Science, Faculty of Medicine, Universitas Brawijaya, Malang, Indonesia ²Department of Radiology, Faculty of Medicine, Universitas Brawijaya, Malang, Indonesia ³Department of Clinical Pathology, Faculty of Medicine, Universitas Brawijaya, Malang, Indonesia ⁴Physics Department, Faculty of Mathematics and Natural Science, Universitas Brawijaya, Malang, Indonesia

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ABSTRACT

Dengue hemorrhagic fever (DHF) is still a major problem in the world. It diagnosis is based on clinical and supporting examinations including leukocytes and platelets examinations, as well as antigen and antibody tests for dengue virus. Prompt and early diagnosis will reduce the risk of disease severity. Examination of lymphocytes and atypical lymphocytes are done to strengthen the diagnosis of dengue hemorrhagic fever patients. Lymphocytes in dengue hemorrhagic fever have their own characteristics and roles in relation to the body's immune system. Lymphoctes and atypical lymphocytes are increased in DHF patients. In this review, the characteristics and role of lymphocytes and atypical lymphocytes are described in dengue hemorrhagic fever.



GRAPHICALABSTRACT

Introduction

Dengue hemorrhagic fever (DHF) can be a lifethreatening disease if it develops into dengue shock syndrome (DSS). The DHF signs include high and sudden fever, thrombocytopenia, leukopenia, lymphocytosis, increased and hematocrit which cause bleeding can complications, severe shock with undetected blood pressure, and pulse which can result in the Diagnosis of DHF is based on the death [1]. World Health Organization (WHO) criteria which are classified into four degrees based on the disease In of DHF, severity. cases thrombocytopenia and leukopenia were found. Neutropenia, eosinopenia, basopenia, monocytopenia, lymphocytosis, and atypical lymphocytes were found changes in color, contour, and texture on a 60x magnification microscope [2].

Lymphocytes in dengue infection

Lymphocytes are white blood cells found in the immune system and mainly play a role in adaptive immunity. Lymphocytes are generally divided into B lymphocytes (B cells), T lymphocytes (T cells), and natural killer cells (NK cells). Lymphocytosis is occurred due to infection dengue [3-5].

In DHF, there is an increase in viral replication in monocytes or macrophages through Fcy receptors by heterotypic antibodies that fail to neutralize the virus through the Fc portion of Ig-G



which forms a viral antibody complex.

Increase in dengue virus infection was first reported in 1977 where IgG antibodies depended on the Ig-G subclass, this phenomenon was observed using polyclonal and monoclonal dengue virus antibodies [6]. Circulating dengue virus binds to specific IgG and forms immune complexes. These immune complexes are found in 48-72% of DHF patients. The role of dengue virus-specific T lymphocytes in the infection healing is not known. It is suspected that dengue virus-specific CD4+ and CD8+ T lymphocytes are able to lyse cells infected with dengue virus by eliminating these monocyte cells. Therefore, controlling infection, virus-specific T lymphocyte activation depends on the viral epitope, especially NS3, E, and NS1 which occupy CD4+ and CD8+16. The dengue virus antibody complex causes cross-reactive activation of CD4+ and CD8+ cytotoxic lymphocytes releasing cytokines and lysing infected monocytes mediated by these lymphocytes [7]. There are several theories about lymphoctes in DHF.

First is the mediator theory, the mediator theory is based on several things, namely virus-infected macrophages secrete mediators or cytokines, the function of cytokines is as mediators in natural immunity caused by stimulation of infectious substances, as regulators regulate lymphocyte activation, proliferation, and differentiation, as a non-specific activator of inflammatory cells and as a stimulator of growth and differentiation of mature leukocytes. The critical period in DHF is short, and then followed by a fast healing period without sequelae, experts have compared it to septic shock [8].

This theory was developed together with the role of endotoxin and lymphocyte cells. Endotoxin will activate the cytokine cascade especially TNF-ά and interleukin-1. In DHF shock, there is 75% endotoxemia, while in those who are not shocked, there is 50%. Tumor Necrosis Factor Alpha increases from the start of the disease course and will decrease after the infection subsides, interleukin-6 increases in DHF with shock [2].

In general, an incoming viral infection will elicit a response from T lymphocytes, and then virusspecific serotype peptides will be carried by MHC class I and presented on the viral surface. The viral epitopes are recognized by CD4+ and CD8+ which then activate Т lymphocytes Т lymphocytes by releasing lymphokines at higher levels. Activated monocytes release factors as a result of interaction with cytotoxic lymphocytes. This cycle produces high levels of lymphokines such as IL-2, monokines such as TNF and chemical mediators such as C3a and C5a in a short time [9].

The next theory is the theory of apoptosis, namely the physiological process of cell death which is a reaction to various stimulants. This process can be divided into two stages, namely damage to the cell nucleus, and then changes in cell shape and cell membrane permeability. As a result of this apoptosis, fragmentation of the cell's DNA will occur, cytoplasmic vacuolization, blebbing and binding of plasma membrane granulations into subcellular DNA containing apoptotic bodies [10].

Cytotoxic T lymphocytes signal protease proteins that induce target cell apoptosis. As a result of DEN virus infection, activated T lymphocytes show high levels of T expression and are highly susceptible to apoptosis. In severe cases of DHF, there is liver damage, Councilman bodies are present. It is possible that this is an apoptotic process in liver cells. Theories that focus on cellular processes override those of immunopathology. According to the experts in this field, viral replication events in macrophages

are also opposed when viral apoptosis occurs and scattered cells are eaten by macrophages or phagocytosed [11-13].

Furthermore, the endotoxin theory indicates that as a result of shock in DHF, ischemia will occur in the tissues and in the intestine, at that time translocation of bacteria occurs so that both the bacteria and their metabolic products, including endotoxins in the intestinal flora, can enter the circulation as a result of endotoxins, can exacerbate the current shock [14-17].

Intestinal bacterial endotoxin plays a role in the severity of clinical symptoms of DHF, because endotoxin activates the cytokine cascade, especially TNF (Tumor Necrotizing Factor) and interleukin-1 (IL-1). In DHF shock, there is 75% endotoxin and in DHF without shock, there is 60%. In severe gastrointestinal bleeding, endotoxin levels were found to be higher than those with mild gastrointestinal bleeding. Intestinal bacterial endotoxins can enter the blood circulation due to the translocation of intestinal bacteria into the blood circulation due to damage to the intestinal lumen [16, 18].

Atypical lymphocytes

Increase of atypical lymphocytes is an increase in blue plasma lymphocytes, namely reactive lymphocytes as an immune response indicating the presence of a virus and can be observed in peripheral blood smears [19-21]. Dengue virus infection causes activation of the immune system, impaired immune response such as inversion of the CD4/CD8 ratio not only interferes with the immune system's ability to clear the virus, but also causes excess production of cytokines that influence T lymphocytes to differentiate into atypical lymphocytes [22, 23].

The number of blue plasma lymphocytes is calculated per 100 leukocytes. Blue plasma lymphocytes are lymphocytes with dark blue cytoplasm and are larger in size [22]. Wide cytoplasm is with fine to very pronounced vacuolization and clear perinuclear areas. The nucleus is located at one edge of the cell, oval, or kidney-shaped [25]. Nuclear chromatin is coarse and sometimes nucleoli are present in the nucleus. In the cytoplasm, there are no

azurophilic granules, the area adjacent to the erythrocytes is not indented and does not turn blue. If a person's blue plasma lymphocyte value reaches \geq 4%, it can be ascertained that the person is infected with the dengue virus. The average number of blue plasma lymphocytes in patients with dengue fever (DF) and dengue hemorrhagic fever (DHF) peaked on the sixth day of illness [25, 26]. Patients with dengue shock syndrome (DSS) have the highest number of blue plasma lymphocytes at the time of shock. The number of blue plasma lymphocytes for each clinical type has a significant difference and higher according to the severity of clinical spectrum. The more severe the immune response that occurs, the more severe the clinical spectrum experienced [2, 22].

Conclusion

DHF is still a health problem and can lead to severity. Lymphocytes and atypical lymphocytes are increased in dengue virus infection and are related to the dynamics of the host's immune system. However, the DHF diagnosis considers clinical and other supporting aspects to reduce the risk of severity and mortality of patients.

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Authors' Contributions

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ORCID

Ahmad Fahrudi Setiawan <u>https://orcid.org/0000-0003-1369-1835</u> Yuyun Yueniwati Prabowowati Wajib <u>https://orcid.org/0000-0002-1395-5557</u> Kusworini Handono <u>https://orcid.org/0000-0001-8473-0235</u> Setyawan Purnomo Sakti <u>https://orcid.org/0000-0001-8473-0235</u>

References

[1]. a) Muller D.A., Depelsenaire A.C., Young P.R., Clinical and Laboratory Diagnosis of Dengue Virus Infection, The Journal of infectious diseases, 2017, **215**:S89 [Crossref], [Google Scholar], [Publisher]; b) Adawara S., Adamu G., Mamza P., Abdulkadir I. In-silico Design of Oxadiazole Hybrids as Potential Inhibitors of Dengue Virus NS2B-NS3 Protease. Advanced Journal of Chemistry, Section A, 2022, 5:118 [Crossref], [Publisher]; c) Adawara S., Adamu G., Mamza P., Abdulkadir, I. Chemoinformatic Design of Phthalazinone Analogues as Novel Dengue Virus NS2B-NS3 Protease Inhibitors with Enhanced Pharmacokinetics. Advanced Journal of Chemistry, Section A, 2022, 5:175 [Crossref], [Publisher]

[2]. Phakhounthong K., Chaovalit P., Jittamala P., Blacksell S.D., Carter M.J., Turner P., Chheng K., Sona S., Kumar V., Day N.P.J., White L.J., Pan-Ngum W., Predicting the severity of dengue fever in children on admission based on clinical features and laboratory indicators: application of classification tree analysis, *BMC pediatrics*, 2018, **18**:109 [Crossref], [Google Scholar], [Publisher]

[3]. Joshi A.A., Gayathri B.R., Swathi K., Correlation of thrombocytopenia with degree of atypical lymphocytosis as a prognostic indicator in dengue, *International Journal of Research in Medical Sciences*, 2017, **5**:1 [Crossref], [Google Scholar], [Publisher]

[4]. Lotfi A.R., Owaysee Osquee H., Investigating factors affecting hospitalization of patients with mucormycosis after contracting covid-19: A Systematic Review, *International Journal of Advanced Biological and Biomedical Research*, 2023, **11**:35 [Google Scholar], [Publisher]

[5]. Muna N.M., Widodo A.D.W., Endraswari P.D., Arfijanto M.V., Correlation between the bacterial and fungal profiles from the clinical specimens with the CD4 counts and the NLR values of HIV/AIDS patients at tertiary referral hospital in Indonesia, *Journal of Medicinal and Chemical Sciences*, 2023, **6**:2111 [Crossref], [Google Scholar], [Publisher]

[6]. a) Nascimento E.J.M., Huleatt J.W., Cordeiro M.T., Castanha P.M.S., George J.K., Grebe E., Welte A., Brown M., Burke D.S., Margues E.T.A., Development of antibody biomarkers of long term and recent dengue virus infection, Journal of Virological Methods, 2018, 257:62 [Crossref], [Google Scholar], [Publisher]; b) Tadayon N., Ramazani A. In silico Analysis of Sars-CoV-2 Main Protease Interactions with Selected Hyoscyamus Niger and Datura Stramonium Compounds for Finding New Antiviral Agents. Chemical Methodologies, 2023, 7:613 [Crossref], [Publisher]; c) Selmi A., Zarei A., Tachoua W., Puschmann H., Teymourinia H., Ramazani A. Synthesis and Structural Analysis of a Novel Stable Quinoline Dicarbamic Acid: X-Ray Single Crystal Structure of (2 - ((4 - ((2 -(Carboxy(methyl)amino)ethoxy)carbonyl)

quinoline-2-yl)oxy) ethyl) (methyl)-carbamic Acid and Molecular Docking Assessments to Test Its Inhibitory Potential against SARS-CoV-2 Main Protease. *Chemical Methodologies*, 2022, **6**:463 [Crossref], [Publisher]

[7]. Tian Y., Grifoni A., Sette A., Weiskopf D., Human T cell response to dengue virus infection, *Frontiers in immunology*, 2019, **10**:2125 [Crossref], [Google Scholar], [Publisher]

[8]. Katzelnick L.C., Gresh L., Halloran M.E., Mercado J.C., Kuan G., Gordon A., Balmaseda A., Harris E., Antibody-dependent enhancement of severe dengue disease in humans, *Science*, 2017, **358**:929 [Crossref], [Google Scholar], [Publisher]

[9]. Silveira G.F., Wowk P.F., Cataneo A.H.D., Dos Santos P.F., Delgobo M., Stimamiglio M.A., Lo Sarzi M., Thomazelli A.P.F.S., Conchon-Costa I., Pavanelli W.R., Antonelli L.R.V., Báfica A., Mansur D.S., Dos Santos C.N.D., Bordignon J., Human T lymphocytes are permissive for dengue virus replication, *Journal of virology*, 2018; **92**:e02181 [Crossref], [Google Scholar], [Publisher]

[10]. Hottz E.D., Bozza F.A., Bozza P.T., Platelets in immune response to virus and immunopathology of viral infections, *Frontiers in medicine*, 2018, 5:121 [Crossref], [Google Scholar], [Publisher]

[11]. John D.V., Lin Y.S., Perng G.C., Biomarkers of severe dengue disease - a review, *Journal of biomedical science*, 2015, **22**:83 [Crossref], [Google Scholar], [Publisher] [12]. Saniathi N.K.E., Rianto B.U.D., Juffrie M., Soetjiningsih S., Dengue hemorrhagic fever: The role of Soluble E-Selektin, Soluble Intra Cellular Adhesion Molecule-1 (Sicam-1) and Soluble Vascular Cellular Adhesion Molecule -1 (Svcam-1) in overweight children, *Bali Medical Journal*, 2018, **8**:183 [Crossref], [Google Scholar], [Publisher]

[13]. Triningrat A.A.M.P., Somia I.K.A., Utama I.M.D.L., Yuliastini N.P.N.S., Wijayati M.P., Handayani A.T., Dengue hemorrhagic fever with severe ocular complication: case series, *Bali Medical Journal*, 2020, **9**:907 [Crossref], [Google Scholar], [Publisher]

[14]. Saniathi N.K.E., Rianto B.U.D., Juffrie M.S., The Effect of overnutrition toward the risk of dengue shock syndrome in pediatric patient: indepth investigation of sVCAM-1 and adiponectin level, *Bali Medical Journal*, 2018, **7**:244 [Crossref], [Google Scholar], [Publisher]

[15]. Sudarmaja I.M., Swastika I.K., Effectiveness of different detergent solutions as larviside for aedesaegypti larvae, *Bali Medical Journal*, 2015, 4:41 [<u>Crossref</u>], [<u>Google Scholar</u>], [<u>Publisher</u>]

[16]. Pinontoan O.R., Sumampouw O.J., Ticoalu J., Nelwan J.E., Musa E.C., Sekeeon J., The variability of temperature, rainfall, humidity and prevalance of dengue fever in Manado City, *Bali Medical Journal*, 2022, **11**:81 [Crossref], [Google Scholar], [Publisher]

[17]. Glasner D.R., Ratnasiri K., Puerta-Guardo H., Espinosa D.A., Beatty P.R., Harris E., Dengue virus NS1 cytokine-independent vascular leak is dependent on endothelial glycocalyx components, *PLoS Pathog*, 2017, **13**:e1006673 [Crossref], [Google Scholar], [Publisher]

[18]. Yong Y.K., Tan H.Y., Jen S.H., Shankar E.M., Natkunam S.K., Sathar J., Manikam R., Sekaran S.D., Aberrant monocyte responses predict and characterize dengue virus infection in individuals with severe disease, *Journal of translational medicine*, 2017, **15**:121 [Crossref], [Google Scholar], [Publisher]

[19]. Utama I.M.G.D.L., Agustini N.M.A., The role of macrophage Migration Inhibitory Factor (MIF) in pediatric dengue infection at Sanglah Hospital, Bali, Indonesia, *Bali Medical Journal*, 2020, **9**:224 [Crossref], [Google Scholar], [Publisher]

[20]. Sudaryanto A., Ainnurriza U.S., Supratman S., Dewi S.K., Mapping the prevalence of dengue fever in sragen regency Indonesia, *Bali Medical Journal*, 2021, **10**:1107 [Crossref], [Google Scholar], [Publisher]

[21]. Yantie N.P.V.K., Gunawijaya E., Suradipa I.W., Gustawan I.W., Asymptomatic Cardiac Rhythm Abnormality in Children with Dengue Virus Infection, *Bali Medical Journal*, 2016; **5**:351 [Crossref], [Google Scholar], [Publisher]

[22]. Joshi A.A., Gayathri B.R., Muneer F., Dynamics of differential count in dengue. *International Journal of Advances in Medicine*, 2018, **5**:1 [Crossref], [Publisher]

[23]. Rampengan N.H., Daud D., Warouw S., Ganda I.J., Albumin globulin ratio in children with dengue virus infection at Prof. Dr. R. D. Kandou Hospital, Manado Indonesia, *Bali Medical Journal*, 2016, **5**:562 [<u>Crossref</u>], [<u>Google Scholar</u>], [<u>Publisher</u>] [24]. Putri D.F., Husna I., Kurniati M., Primadiamanti A., Activity of enzyme Esterase, Glutathione S Transferase and Inorganic Substance of Dengue vector Aedes aegypti larvae against Lansium domesticum leave extract and fractionation, *Bali Medical Journal*, 2023, **12**:1163 [Crossref], [Google Scholar], [Publisher]

[25]. Pranata I.W.A., Diana A., Heryanto M.R., Lukman N., Kosasih H., Djauhari H., Butarbutar D.P., Widjaja S., Alisjahbana B., Persistence of anti-Salmonella O9 IgM as measured by Tubex® TF may contribute to the over-diagnosis of typhoid fever in endemic areas, *Bali Medical Journal*, 2022, **11**:11 [Crossref], [Google Scholar], [Publisher]

[26]. Novena O.D., Skin manifestations in coronavirus disease 2019 (COVID-19): A literature review, *Indonesia Journal of Biomedical Science*, 2021, **15**:113 [Crossref], [Google Scholar], [Publisher]

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